

Project-based Training Course (280995)
International Program of Frontier Biotechnology
Autumn semester 2010

List of laboratories :

1. Fujiyama Laboratory (2 students)

Supervisor : Assist. Prof. Takao Ohashi (ohashi@icb.osaka-u.ac.jp)

Students : Topic 1 – Poonperm Rawin (Rawin), Topic 2 – Permanasari Etin Diah (Etin)

Topic 1 : Analysis of localization of glycosyltransferase from the fission yeast

Schedule :

Day 1 : Transformation of green fluorescent protein (GFP)-fused glycosyltransferase into the fission yeast

Day 2 : Picking up the transformants

Day 3 : Cultivation of the transformants

Day 4 : SDS-PAGE and observation by fluorescence microscopy

Day 5 : Western blotting

Topic 2 : Quantification of lactic acid produced by *Lactobacillus* using lactose dehydrogenase

Schedule :

Day 1 : Preparation of the buffers for enzymatic reaction

Day 2 : Examine the effects for reaction time and inoculation of the *Lactobacillus* into the milk

Day 3 : Examine the effects for the amounts of the enzyme and lactic acid concentrations

Day 4 : Examine the effects for pH

Day 5 : Quantification of lactic acid produced by *Lactobacillus*

2. Fukui Laboratory (2 students)

Supervisor : Assoc. Prof. Sachihito Matsunaga (sachi@bio.eng.osaka-u.ac.jp)

Student : Ariyasrijit Chananya (Belle)

Topic : Functional analyses of ASURA

Schedule :

Day 1 : Preparation of HeLa cells

Day 2 : Preparation of cell lysates

Day 3 : Western Blotting

Day 4 : Immunostaining

Day 5 : Data analyses

3. Fukusaki Laboratory (2 students)

Supervisor : Assist. Prof. Hisayo Ono (ono@bio.eng.osaka-u.ac.jp)

Students : Topic 1 - Peerakietkhajorn Saranya (Tip), Topic 2 – Jung Jieun (Jieun)

Topic 1 : Metabolic profiling of different tea leave products, green tea, black tea, and oolong tea, based on gas chromatography/mass spectrometry (GC/MS)

Schedule :

Day 1 : Extraction of metabolites of tea leaves

Day 2 : Derivatization and GC/MS analysis

Day 3 : Data preprocessing for multivariate analysis

Day 4 : Data analysis (multivariate analysis): Principle component analysis

Day 5 : Data analysis (metabolite identification)

Topic 2 : Metabolic profiling of ranked Japanese green tea based on gas chromatography/mass spectrometry (GC/MS)

Schedule :

Day 1 : Extraction of metabolites of tea leaves

Day 2 : Derivatization and GC/MS analysis

Day 3 : Data preprocessing for multivariate analysis

Day 4 : Data analysis (multivariate analysis): Principle component analysis

Day 5 : Data analysis (metabolite identification)

4. Harashima Laboratory (2 students)

Supervisor : Assoc. Prof. Yoshinobu Kaneko (kaneko@bio.eng.osaka-u.ac.jp)

Students : Topic 1 – Suandi Pratama Sultan (Andi), Topic 2 – Ly Nhung Hong (Nhung)

Topic 1 : Isolation of yeast auxotrophic mutant by UV mutagenesis

Schedule :

Day 1 : Preparation of media and inoculation of wild-type strain

Day 2 : UV irradiation

Day 3 : Incubation

Day 4 : Replica plating

Day 5 : Judgement of auxotrophy

Topic 2 : Knockout yeast gene

Schedule :

Day 1 : Preparation of media and inoculation of host strain

Day 2 : Preparation of disruption cassette by PCR

Day 3 : Transformation

Day 4 : Incubation

Day 5 : Picking up candidate and check by colony PCR

5. Kanaya Laboratory (2 students)

Supervisor : S. A. Assist. Prof. Eiko Kanaya (ekanaya@mls.eng.osaka-u.ac.jp)

Student : Topic 1 – Salayev Ahmed (Ahmed), Topic 2 – no one

Topic 1 : Production and thermal denaturation of Tma-RNase H1

Schedule :

Day 1 : Media preparation and pre-culture of Tma-RNase H1

Day 2 : Overproduction of Tma-RNase H1

Day 3 : Cell harvest, cell disruption, cell supernatant separation, dialysis, SDS-PAGE preparation

Day 4 : Purification by cation and affinity column chromatography and SDS-PAGE analysis

Day 5 : CD spectra and thermal denaturation of Tma-RNase H1

Topic 2 : Chaperone function of SIB1 FKBP22, a peptidyl prolyl cis-trans isomerase from psychrotrophic bacterium

Schedule :

Day 1 : Overproduction SIB1 FKBP22

Day 2 : Purification 1: Affinity chromatography (Ni²⁺ column); dialysis; gel filtration wash

Day 3 : Purification 2: Gel filtration, dialysis

Day 4 : Chaperone function 1 : binding affinity to a folding intermediate protein
Day 5 : Chaperone function 2 : aggregation prevention

6. Kino-oka Laboratory (1 student)

Supervisor : Assist. Prof. Mee-Hae Kim (mh-kim@bio.eng.osaka-u.ac.jp)

Student : no one

Topic : Understanding the animal cell culture and characterization of cell properties

Schedule :

Day1 : Culture of human fibroblasts

Day2 : Taking cell pictures and counting number of adherent cells

Day3 : Changing medium

Day4 : Staining of F-actin and observation by fluorescence microscopy

Day5 : Image and data analysis: cell attachment, morphology, and proliferation

7. Muranaka Laboratory (1 student)

Supervisor : Assist. Prof. Atsushi Okazawa (okazawa@bio.eng.osaka-u.ac.jp)

Student : Limkul Juthamard (Jah)

Topic : Cloning of cellulase from parasitic plant, Orobanche minor

Schedule :

Day 1 : cDNA preparation from haustoria of *O. minor*

Day 2 : PCR of partial sequences of cellulase genes

Day 3 : RACE of the genes

Day 4 : Cloning of full length genes

Day 5 : Transformation (*E. coli*)

8. Nihira Laboratory (2 students)

Supervisor : Assist. Prof. Shigeru Kitani (kitani@icb.osaka-u.ac.jp)

Students : Maria Odaise Silva dos Santos (Daise), Jumhawan Udi (Udi)

Topic : Measurement and detection of microbial secondary metabolites

Schedule :

Day 1 : Preparation of medium and inoculation

Day 2 : Bioassay

Day 3 : Extraction with organic solvent and evaporation

Day 4 : HPLC detection

Day 5 : TLC detection

9. Watanabe Hajime Laboratory (2 students)

Supervisor : Assist. Prof. Koji Tsukada (tsukada_koji@bio.eng.osaka-u.ac.jp)

Students : Hong Xun(Hong), Natesuntorn Waranya (Sine)

Topic : Study of the anti-virus effect of silver (Ag-Zeolyte) which was known as a bacteriostatic substance and electron microscopic study of natural coliphage species around us.

Schedule :

Day 1 : Preparation of culture medium, agar plate, *E. coli* cells, and RNA phage. Sampling and partial purification of some phage samples (natural water) in our university.

Day 2 : Training of basic procedures of virus research : Plaque assay, one-step growth experiment and single burst experiment.

- Day 3 : Evaluation of Ag-zeolyte effect on the bacterial growth, the infectivity of RNA coliphage Q β , and some natural phages.
- Day 4 : Preparation of specimens for electron microscopic observation of phages and phage-infected cells.
- Day 5 : Transmission electron microscopy observation of the shape of phage particles and infected bacterial cells by using a negative staining.